

# HISTOLOGY SAMPLE COLLECTION & SHIPPING INSTRUCTIONS



Thank you for submitting samples to the Centre for Crocodile Research ("CCR").

Please carefully follow the instructions below for correctly collecting samples for histology. Samples collected in deviation from this format may result in inaccurate and/or unusable results. Sample collection and shipping is the responsibility of the Client.

Please ensure:

- all samples are collected by suitably qualified personnel in compliance with all regulatory guidelines and ethical codes;
- samples are uniquely labelled for identification with a permanent water-resistant marker and listed on the "Sample Lodgement Form" (page 4) to be printed and included with the samples as well as emailed to [admin@crocresearch.com.au](mailto:admin@crocresearch.com.au);
- a copy of your CITES export permit is provided as soon as possible so that CCR can apply for their CITES import permit; and
- most importantly, please email us ([admin@crocresearch.com.au](mailto:admin@crocresearch.com.au)) if you need any assistance with sample collection, documentation, etc. We will be more than happy to assist you and make this process as easy as possible.

Kind regards,

**Sally Isberg**

Director, Centre for Crocodile Research

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## Sample collection instructions for crocodile skin histology samples

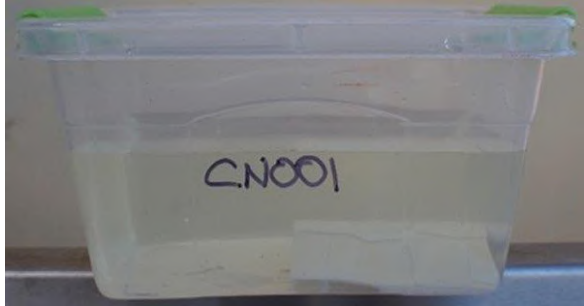

CITES import/export permits will be required for these samples. Before CCR can apply for an import permit, you will need to obtain an export permit from your CITES authority. Once the export permit is provided to CCR, the Australian import permit will be submitted and can take 40 business days before being issued.

Fixing crocodile skin in formalin can change certain characteristics so to ensure the correct lesion/s are sampled, it is necessary to document the sample as follows.

<p>1. Take a photo of the scale(s)/lesion(s) of interest (both with and without light table if necessary). Ensure a ruler is in the photo for reference. Label photos with unique sample ID and suffix. Eg, CN001_without light-table CN001_with light-table</p>	<p>Without light table      With light table</p>
<p>2. Cut the scale/s out with a 1cm border and label with unique ID (in <b>pencil ONLY</b>). Clearly mark lesion of interest with an arrow in <b>pencil ONLY</b>. Re-take photo to show exact boundaries and label with sample ID and suffix (Eg CN001_marked).</p>	
<p>3. Describe the lesion on the Sample Lodgement Form against the unique ID. Please include as much detail as possible. This form must accompany the shipment as well as be emailed to CCR (<a href="mailto:admin@crocresearch.com.au">admin@crocresearch.com.au</a>). Photos should also be emailed.</p>	

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<p>4. Place sample in 10 times the volume of 10% formalin for <u>72 hours only</u>. Label container.</p> <p>For <b>skin</b>, please ensure the container is large enough that the skin lays <u>flat</u>.</p>	
<p>5. After <u>72 hours</u>, drain formalin and replace with 70% alcohol. Leave until shipment.</p>	
<p>6. On the day of shipment, remove sample from container and wrap in 70% alcohol soaked paper-towel. If skin, ensure it lays flat. Seal into labelled zip-lock bag. Keep cool but does not require refrigeration.</p>	

Please note that 70% ethanol is the preferred alcohol but 70% methylated spirits is also acceptable.

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	SampleID	Scale location	Photos			Description of lesions/problem
			No light table	Light table	Marked	
1			Y / N	Y / N	Y / N	
2			Y / N	Y / N	Y / N	
3			Y / N	Y / N	Y / N	
4			Y / N	Y / N	Y / N	
5			Y / N	Y / N	Y / N	
6			Y / N	Y / N	Y / N	
7			Y / N	Y / N	Y / N	
8			Y / N	Y / N	Y / N	
9			Y / N	Y / N	Y / N	
10			Y / N	Y / N	Y / N	
11			Y / N	Y / N	Y / N	
12			Y / N	Y / N	Y / N	
13			Y / N	Y / N	Y / N	
14			Y / N	Y / N	Y / N	
15			Y / N	Y / N	Y / N	
16			Y / N	Y / N	Y / N	
17			Y / N	Y / N	Y / N	
18			Y / N	Y / N	Y / N	
19			Y / N	Y / N	Y / N	
20			Y / N	Y / N	Y / N	